

Optimization of Cardiac Myocyte Model for CiPA Initiative

Zhihua Li, Ph.D.

Division of Applied Regulatory Science
Office of Clinical Pharmacology, Office of Translational Sciences
Center for Drug Evaluation and Research
U.S. Food and Drug Administration

Cardiac Physiome CiPA meeting 2017
Nov 2017



Disclaimer

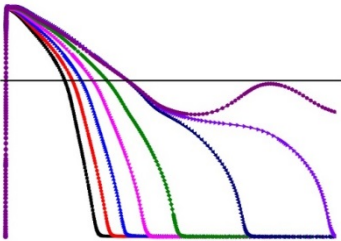
This presentation is not an official US Food and Drug Administration guidance or policy statement. No official support or endorsement by the US FDA is intended or should be inferred.

In Silico Working Group

Goals: Integrate in vitro data into a computational model of human ventricular myocyte and identify a mechanistic metric that can quantify the relative risk of inducing EAD/TdP

In silico Reconstruction of Human Ventricular Cardiomyocyte Electrophysiology

$$I_{\text{stim}} = C \frac{dV_m}{dt} + I_m$$



- Select a consensus base cardiomyocyte model for CiPA
- The base model is to be further optimized based on experimental data of drug effects on selected human cardiac currents
- A set of 12 training drugs classified into 3 torsade de pointes (TdP) risk categories (high, intermediate and low) is used to calibrate the model and develop the metric; Another set of 16 drugs for independent validation

The Selection of the Base Model for CiPA

- Cardiac Modeling Experts Meeting (July 2013) held at FDA to kick off CiPA In Silico Workstream
 - Modeling experts from academics, industry, FDA – hosted by HESI
- Affirmed the use of single cell vs. more complex 2D or 3D models
 - Simple but experimentally determined mechanistic representation of electrophysiology and pharmacology
 - Concerns about the degree of uncertainty generated by the large number of free parameters in a more complex model
 - Interest in quantitative metric(s) that could assign a level of risk vs. simulating proarrhythmia
- Recommended O’Hara Rudy (ORd) human ventricular myocyte model as most tightly linked to human ventricular cell data
- Identified the probable need to consider dynamic drug-channel interactions for hERG (and other channels)

Improving the ORd Model for CiPA

- Making the IKr/hERG component temperature dependent
- Modeling dynamic drug-hERG interactions rather than using simple IC50s
- Optimizing model parameters based on experimentally recorded drug effects on human ventricular myocytes

Development of a Temperature Sensitive hERG Model

FDA



Contents lists available at [ScienceDirect](https://www.sciencedirect.com)

Journal of Pharmacological and Toxicological Methods

journal homepage: www.elsevier.com/locate/jpharmtox



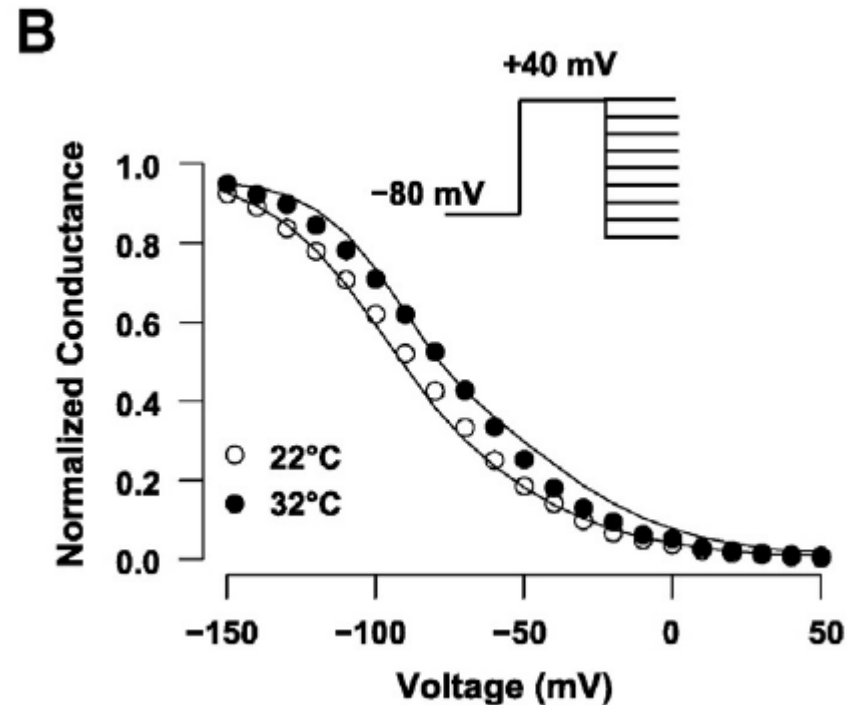
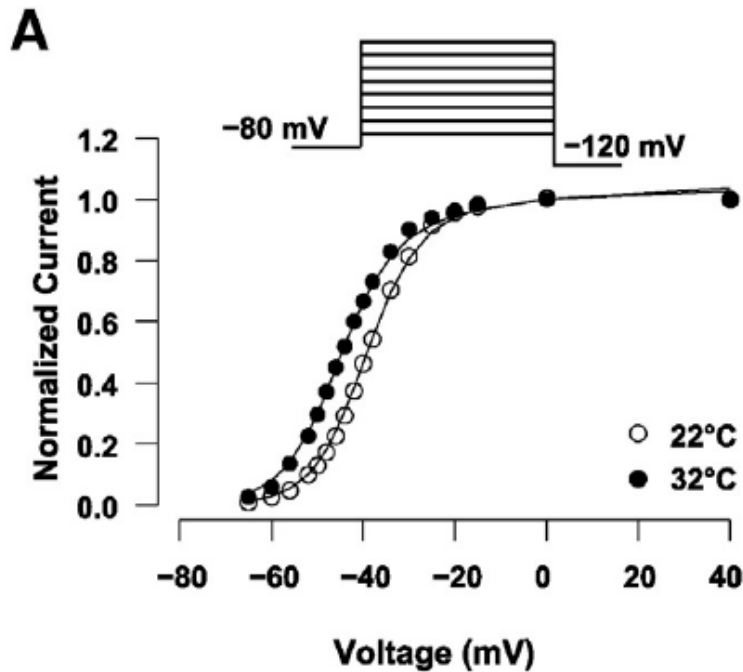
A temperature-dependent *in silico* model of the human ether-à-go-go-related (hERG) gene channel

Zhihua Li *, Sara Dutta, Jiansong Sheng, Phu N. Tran, Wendy Wu, Thomas Colatsky

Division of Applied Regulatory Science, Office of Clinical Pharmacology, Office of Translational Sciences, Center for Drug Evaluation and Research, Food and Drug Administration, United States

- Because O'Hara-Rudy model operates at physiological temperature, while industry-generated hERG data are often obtained at room temperature, a dynamic, temperature-sensitive hERG model is required
- We developed a modified hERG model that can reproduce temperature-induced changes in major channel gating processes

Examples of Temperature Effects



Experimental data (circles) from Vandenberg et al. 2006; Simulation (lines) from Li et al. 2016.

- Our model was able to reproduce the experimentally observed left shift of steady state activation curve (A)
- And a right shift of steady state inactivation curve (B)

Original Article

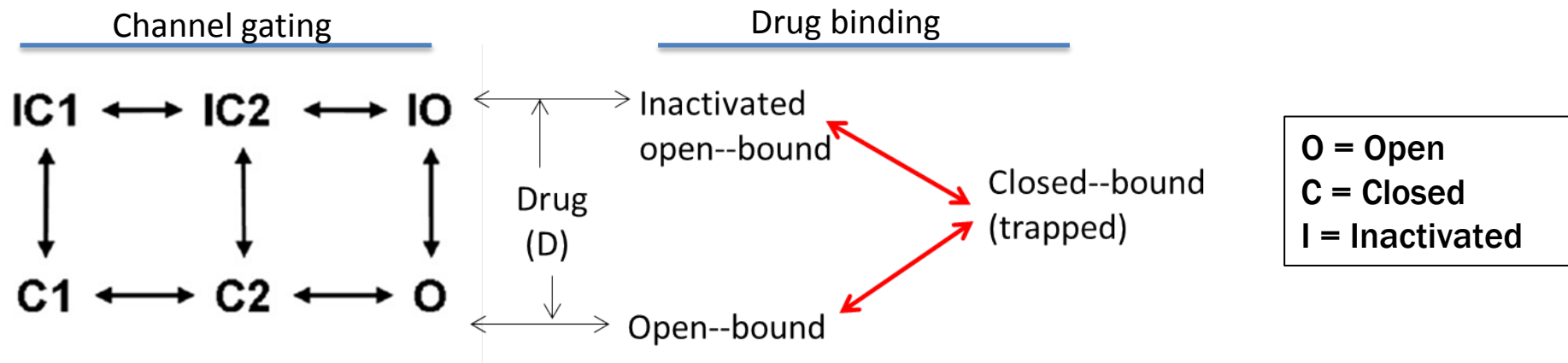
Improving the In Silico Assessment of Proarrhythmia Risk by Combining hERG (Human Ether-à-go-go-Related Gene) Channel-Drug Binding Kinetics and Multichannel Pharmacology

Zhihua Li, PhD; Sara Dutta, PhD; Jiansong Sheng, PhD; Phu N. Tran, PhD; Wendy Wu, PhD; Kelly Chang, PhD; Thembi Mdluli, PhD; David G. Strauss, PhD; Thomas Colatsky, PhD

Li Z et al. Improving the In Silico Assessment of Proarrhythmia Risk by Combining hERG-Drug Binding Kinetics and Multichannel Pharmacology. *Circulation: Arrhythmia & Electrophysiology*. 2017;10:e004628

- Because the same drug may show different block potency under different conditions (i.e. heart rate), a novel model was developed to capture this dynamic drug-hERG interaction
- This model can distinguish between hERG blockers with similar IC50s but different TdP liabilities because of some drugs' tendency to be trapped in closed hERG channel

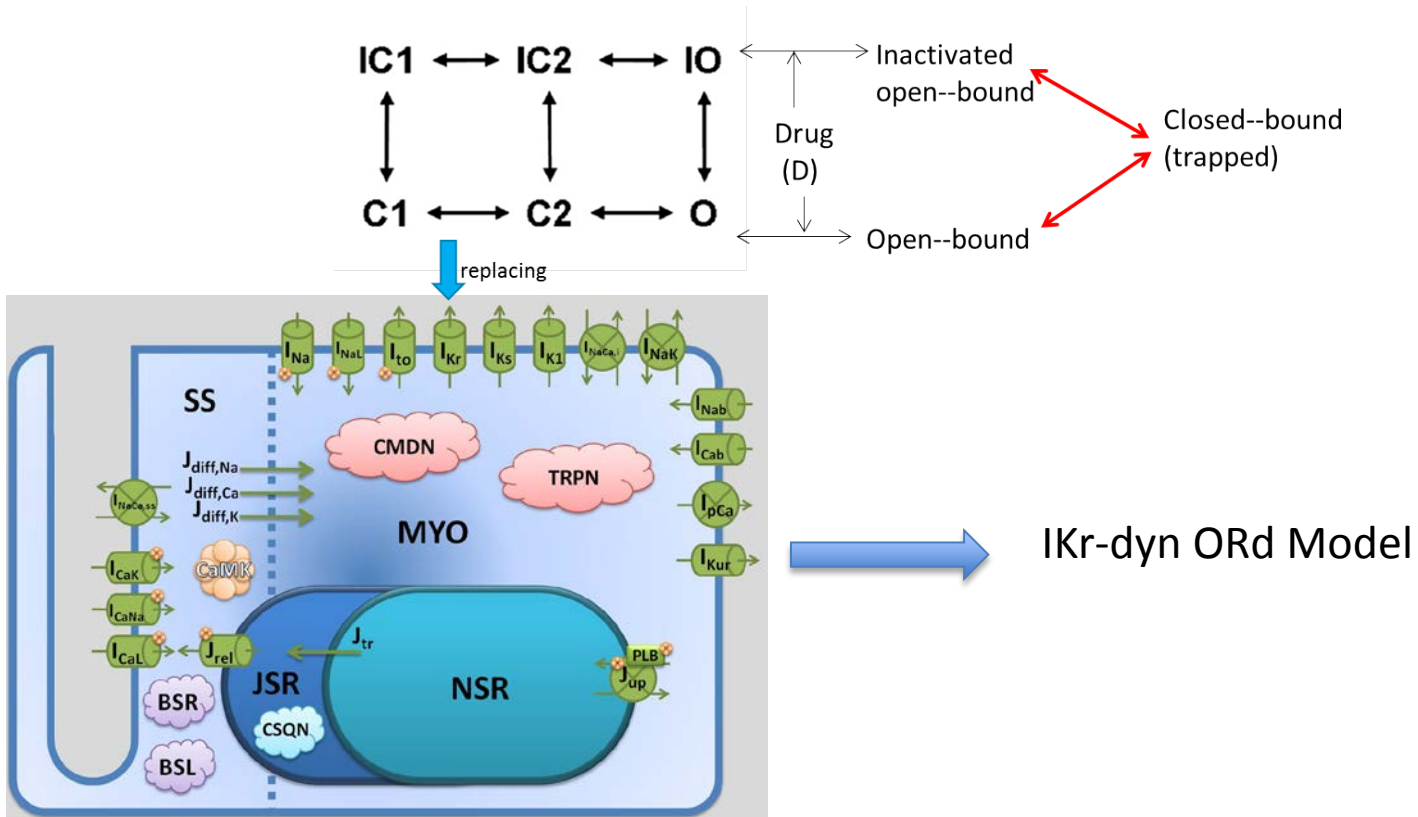
Modeling Dynamic drug-hERG Interactions



Li Z et al. Circulation: Arrhythmia & Electrophysiology. 2017;10:e004628

- The model allows drugs to be trapped in closed-bound state with varying propensities, a realistic feature often missing from published hERG models
- Modeling shows that High TdP Risk compounds tend to have a higher propensity to be trapped within hERG channel during repolarization

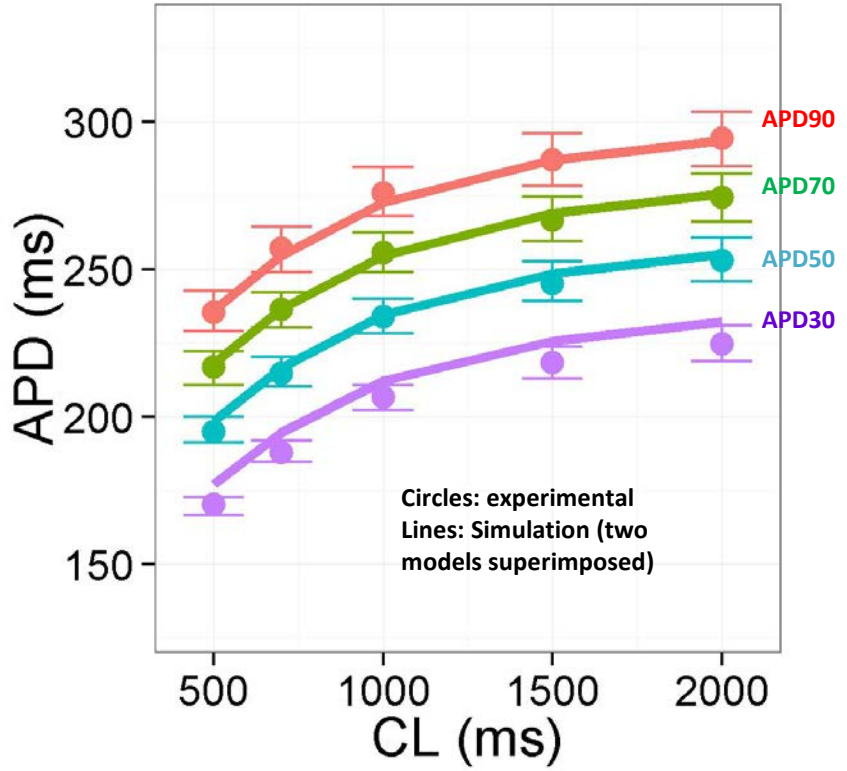
Replacing the IKr component of ORd with the Dynamic hERG Model



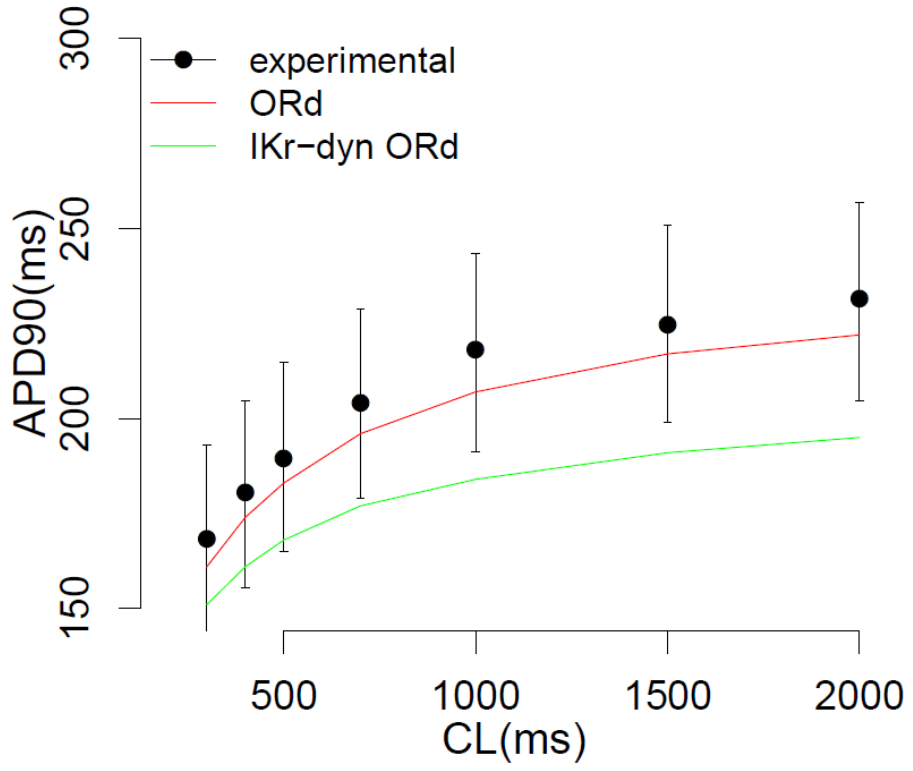
O'Hara T, Virag L, Varro A, & Rudy Y (2011) *PLoS Comput Biol* 7(5):e1002061.

IKr-dyn ORd Model vs ORd Model

Control Condition (no drug)



1 μ M nisoldipine (ICaL blocker)



- Under control conditions, both models fit experimental rate-dependent APD well
- In the presence of some drugs, IKr-dyn ORd model made the fitting worse

Further Optimization of IKr-dyn ORd



Optimization of an *In silico* Cardiac Cell Model for Proarrhythmia Risk Assessment

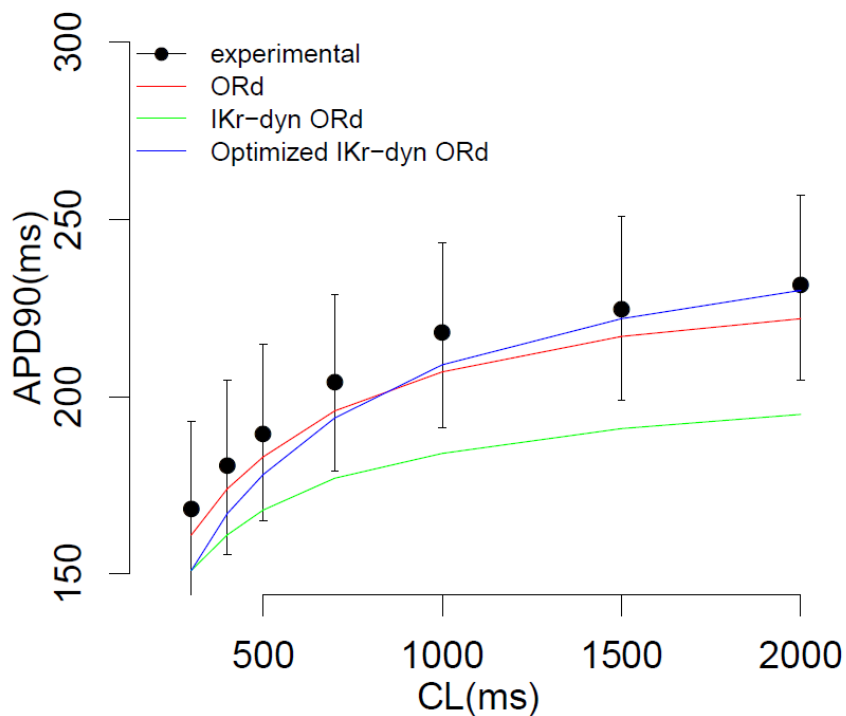
Sara Dutta, Kelly C. Chang, Kylie A. Beattie, Jiansong Sheng, Phu N. Tran, Wendy W. Wu, Min Wu, David G. Strauss, Thomas Colatsky[†] and Zhihua Li^{*}

Division of Applied Regulatory Science, Office of Clinical Pharmacology, Office of Translational Sciences, Center for Drug Evaluation and Research, U.S. Food and Drug Administration, Silver Spring, MD, United States

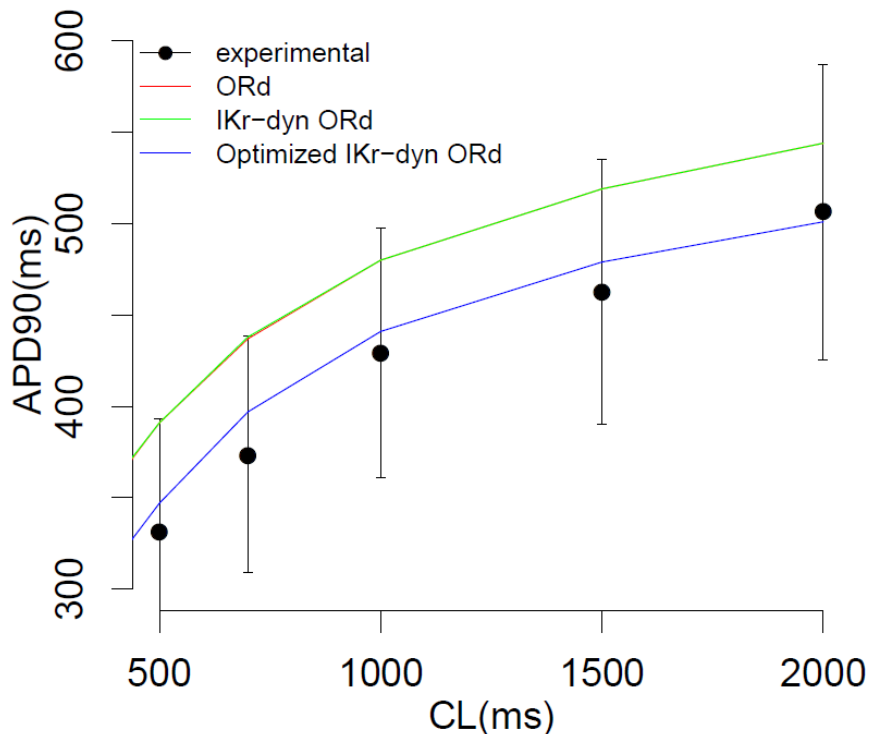
- The conductance of five major ion currents (IKr, IK1, IKs, INaL, ICaL) are adjusted
- Adjustment is based on APD rate dependence experimental data under control and drug block conditions in human cardiomyocytes (from O’Hara 2011)

Results of Optimization

1 μ M nisoldipine (ICaL blocker)



1 μ M E4031 (IKr blocker)



- For some drugs (nisoldipine), Optimized IKr-dyn ORd model (blue) reproduces data as faithfully as ORd (red)
- For others (E4031), Optimized IKr-dyn ORd model (blue) fits experimental data even better than ORd

Summary



- The consensus cardiac model (ORd) was selected based on its tight link to human ventricular cell data
- The choice of model complexity (1D vs 2D) and tissue/cell type is based on the assumption that the goal is to assign relative TdP risk, not to simulate TdP directly
- IKr component of ORd was replaced by a Markov hERG model that captures temperature-dependent gating and drug-hERG dynamic interaction
- Further optimization of the model improves the model's ability to reproduce rate-dependent APD experimental data from human cardiomyocytes



Acknowledgements

CiPA Steering Committee

Ayako Takei, Bernard Fermini, Colette Strnadova, David Strauss, Derek Leishman, Gary Gintant, Jean-Pierre Valentin, Jennifer Pierson, Kaori Shinagawa, Krishna Prasad, Kyle Kolaja, Natalia Trayanova, Norman Stockbridge, Philip Sager, Tom Colatsky, Yuko Sekino, Zhihua Li, Gary Mirams

All CiPA Working groups

- Ion Channel working group
- In silico working group
- Cardiomyocyte working group
- Phase 1 ECG working group

ALL contributors to CiPA (there are a lot!)

- Public-private partnerships: HESI, SPS, CSRC
- Regulatory Agencies: FDA, EMA, PMDA/NIHS, Health Canada
- Many pharmaceutical, CRO, and laboratory device companies
- Academic collaborators

FDA Contributors

- Norman Stockbridge
- Christine Garnett
- John Koerner

In silico / ion channel

- Zhihua Li
- Wendy Wu
- Sara Dutta
- Phu Tran
- Jiangsong Sheng
- Kelly Chang
- Kylie Beattie
- Min Wu
- Richard Gray

Cardiomyocyte

- Ksenia Blinova
- Derek Schocken
- Li Pang

Phase 1 ECG biomarker

- Jose Vicente
- Lars Johannesen
- Meisam Hosseini
- Alexander Wong
- Dustin McAfee
- Robbert Zusterzeel
- Krystal Lansdowne

